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Original Research Article (Experimental)

Antitumor effect of *Ferula assa foetida* oleo gum resin against breast cancer induced by 4T1 cells in BALB/c miceSeyyed Majid Bagheri <sup>a, b</sup>, Amir Abdian-Asl <sup>c, \*</sup>, Mahin Taheri Moghadam <sup>d</sup>, Maryam Yadegari <sup>e</sup>, Aghdas Mirjalili <sup>e</sup>, Fatemeh Zare-Mohazabieh <sup>f</sup>, Haniyeh Momeni <sup>f</sup><sup>a</sup> Dep. of Physiology, Shahid Sadoughi University of Medical Sciences, Yazd, Iran<sup>b</sup> Neurobiomedical Research Center, Shahid Sadoughi University of Medical Sciences, Yazd, Iran<sup>c</sup> Dep. of Immunology, Shahid Sadoughi University of Medical Sciences, Yazd, Iran<sup>d</sup> Dep. Anatomical Sciences, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran<sup>e</sup> Dep. Anatomical Sciences, Shahid Sadoughi University of Medical Sciences, Yazd, Iran<sup>f</sup> School of Medicine, Shahid Sadoughi University of Medical Sciences, Yazd, Iran

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## ABSTRACT

**Background:** *Ferula assa foetida* commonly consumed as a healthy beverage has been demonstrated to have various biological activities, including antioxidation, anti-obesity and anti-cancer.**Objective:** Our study aims to investigate the antitumor effect of asafoetida *in vivo* using mouse mammary carcinoma 4T1 cells.**Materials and methods:** In the study, female BALB/c mice were divided into two groups (n = 6), which were control (untreated) and other group of mice with breast cancer treated with 100 mg/kg of asafoetida, respectively, by oral gavage. All mice were injected into the mammary fat pad with 4T1 cells (1 × 10<sup>5</sup> 4T1 cells/0.1 ml of phosphate buffer solution). Asafoetida was administered on day 15 after the tumor had developed for 3 weeks. At end of experiment, tumor weight, tumor volume and tumor burden were measured and lung, liver, kidney and tumor were harvested and sections were prepared for histopathological analysis. Lipoxygenase inhibitory and antioxidant activity of asafoetida was also determined.**Results:** Our results showed that treatment with asafoetida was effective in decreasing the tumor weight and tumor volume in treated mice. Body weight significantly increased in female BALB/c mice against control. Apart from the antitumor effect, asafoetida decreased lung, liver and kidney metastasis and also increased areas of necrosis in the tumor tissue respectively.**Conclusions:** The present study demonstrated that asafoetida has potent antitumor and antimetastasis effects on breast cancer and is a potential source of natural antitumor products.© 2017 Transdisciplinary University, Bangalore and World Ayurveda Foundation. Publishing Services by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

Cancer is a major public health problem in many parts of the world. It is currently the second leading cause of death and is expected to surpass heart diseases as the leading cause of death in the next few years [1]. In industrialized countries, breast

cancer is the most common type of cancer among women [2]. Despite the advance of cancer prevention methods early diagnostic detection and used of anti-estrogenic drugs such as tamoxifen and raloxifene, breast cancer is still the most commonly diagnosed cancer that contributes to the second highest cause of cancer associated mortality in women [3]. The conventional therapies for cancer include chemo- and radiotherapies mediated by inducing apoptosis or inhibiting proliferation in neoplastic cells [4]. These therapies cause damage to healthy tissues around the tumors [5] and also develop resistance by numerous tumors [6]. Researchers have been studying alternatives of cancer therapy by applying potential biological

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molecules to target neoplastic tumors [7]. Herbs have been identified as an important source of novel bioactive compounds for medicine development including cancer chemotherapeutic drugs [8]. The efficacy of these herbal chemopreventive agents has been related with their antioxidant potential to reduce or inhibit free radical mediated damage to cellular macromolecules, such as DNA, lipids and proteins [9]. *Ferula assa-foetida* L. is one of these species that grows wild in central area of Iran. The part used of this plant and several other species of *Ferula* is an oleo gum resin [asafoetida] that obtained by incision of stem and root [10]. In Iranian folk medicine, asafoetida is considered as an antispasmodic, anthelmintic, sedative, anticonvulsant and carminative agent [11]. Recent pharmacological and biological studies have verified several pharmacological activities such as antioxidant [12], antileishmanial [13], anticonvulsant [14] anti-diabetic [15], antispasmodic [16], hypotensive [17], and antinociceptive [18] from this oleo gum resin. A meta-analysis study showed that in countries like Japan, Russia, China, Indonesia, the rate of cancer is higher in the comparison of countries that usage of asafoetida is common [19]. Recent studies have revealed its potential antioxidant, antimutagenicity and cancer chemopreventive activities [11]. Although some evidences has been shown that asafoetida could act as anticarcinogen, to our knowledge, no studies are available demonstrating the breast cancer chemopreventive efficacy of *F. assa foetida* oleo gum resin using an in vivo model of breast cancer. This study provides the scientific evidence for the chemopreventive potential of asafoetida against breast cancer induced by 4T1 cells in BALB/c mice.

## 2. Materials and methods

### 2.1. Plant oleo-gum resin

*F. assa foetida* oleo-gum-resin was collected during the summer from Tabas region (Yazd province, Iran) during the summer and the plant species was botanically identified by Dr. Abbas Zarezadeh in Yazd Agricultural Research Center. The dried powder of asafoetida was soaked in distilled water overnight at room temperature and the yielded suspension was used orally. Concentrations and dosages of the extract were expressed as crude amount of dried oleo-gum-resin used in preparing the stock solution [20].

### 2.2. Animals and treatment

8-week old female BALB/c mice bred in animal house of Medical School of Shahid Sadoughi University of Medical Sciences were selected. The mice were housed in 12 h of light and dark cycle and fed with standard pellet and water *ad libitum*. 10 mice were inoculated with  $1 \times 10^5$  4T1 cells/mice on left mammary fat pad subcutaneously. 2 weeks after cancer cell implantation, mice were randomly assigned into two groups: control group (normal saline) and asafoetida group (100 mg/kg BW, orally fed every day). Asafoetida treatment was continued until day 21 post-inoculation. At the end of the experiment, mice were sacrificed and tumor, lung, liver and kidney were removed for histopathological studies [21].

### 2.3. Measurement of body weight and tumor sizes

Body weight was measured every five days. Tumors sizes (tumor weight, tumor volume and tumor burden) was calculated at the end of experiment using the following formula: tumor volume ( $\text{mm}^3$ ) = [(width)<sup>2</sup> × length]/2 and tumor burden (%) = tumor volume ( $\text{mm}^3$ )/body weight (g) × 100 [21].

### 2.4. Histopathological studies

Lungs, liver, kidney and tumor were harvested, fixed in 10% formaldehyde (Sigma–Aldrich, USA) and embedded in paraffin, cut into 5- $\mu\text{m}$  sections, and stained with hematoxylin and eosin (H&E). Sections were evaluated for tumor cell cytology, mitotic rate, growth pattern, necrosis, and metastatic tumor nodules present on these tissues.

### 2.5. Lipoxygenase inhibition activity of asafoetida

The soybean 15-lipoxygenase was used to test the 15-LOX inhibitory activity of asafoetida. For this purpose, 50 ml of extract solution was added to test solution containing: 3 ml of phosphate buffer (0.1 M, pH = 8), 50 ml enzyme solution (final concentration: 167 U/ml) to achieve the enzyme inhibition between 20 and 80%. After 4 min incubation of test solution, the substrate (Linoleic acid, final concentration: 134 mM) was added and the change in absorbance was measured for 60 s at 234 nm. The IC<sub>50</sub> value was calculated graphically using the slopes of absorbance curves. The enzyme solution was kept in ice and tested at intervals to ensure that the enzyme activity was constant. All experiments were performed by UV/Vis Unicco Double Beam Spectrophotometer at 25 °C in triplicate [22].

### 2.6. Antioxidant activity assay of asafoetida

To determine the ability of asafoetida to scavenge DPPH radicals, freshly prepared ethanolic DPPH (0.1 mM; 1 ml) solution was added to different concentrations of asafoetida (20–200  $\mu\text{g}/\text{ml}$ ). After half an hour, the absorbance was recorded at 517 nm. Results were expressed as percentage inhibition as:

$$\text{Inhibition(\%)} = [(A \text{ Control} - A \text{ Extract})/A \text{ Control}] \times 100$$

Where A Control is the absorbance of a DPPH solution without asafoetida, A Extract is the absorbance of the tested extract, which is equal to the absorbance of the asafoetida plus the DPPH (20 mg/L) minus the blank extract absorbance. The samples were run in triplicate and the mean value of three of them was recorded. The percentage inhibition was plotted against the sample concentration in order to calculate the IC<sub>50</sub> values, which is the concentration ( $\mu\text{g}/\text{ml}$ ) of asafoetida that causes 50% loss of DPPH activity [12].

### 2.7. Statistical analysis

Differences in the experiments were analyzed for statistical significance by Student's *t*-test (two-tailed). Results were expressed as mean ± standard error (SEM). Values of  $p < 0.05$  were considered to be statistically significant.

## 3. Results

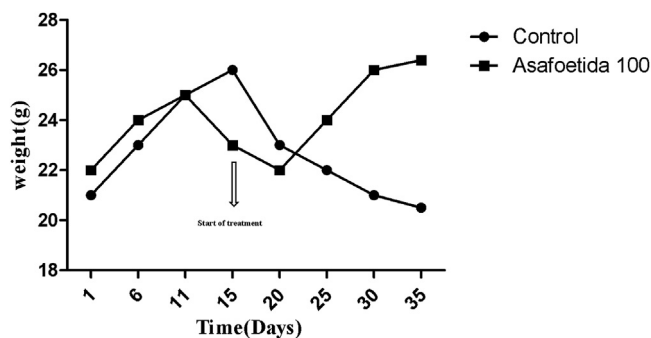
### 3.1. Measurement of body weight and tumor sizes

At the end of the experiment, tumors were excised from each animal for examination of tumor weight and assessment of the antitumor effect of asafoetida. Tumor weight, tumor volume and tumor burden decrease significantly in the asafoetida-treated group when compared with control (Table 1). Changes in the body weight of animals were studied during the experimental period of 5 weeks. It was observed that significant body weight loss was found in control group. After asafoetida administration, the body weight started to increase suggesting that asafoetida treatment ameliorated damage to overall body metabolism (Fig. 1).

**Table 1**  
Effect of asafotida on tumor sizes in animals treated and compared to control group.

Groups	Tumor volume (mm <sup>3</sup> )	Tumor weight (g)	Tumor burden (%)
Control	1512 ± 125	2.3 ± 0.6	74.2 ± 9.2
Asafotida100	498 ± 44**	1.2 ± 0.2*	18.8 ± 4.5**

Values represent the mean ± SEM, n = 5. \*p < 0.05 and \*\*p < 0.01.



**Fig. 1.** Change in body weight of mice in control group and after treatment with asafotida.

### 3.2. Microscopic study of breast cancer cells

The histology sections of breast showed infiltration of tumor cells. In neoplastic cells large hyperchromatic nuclei and relatively small amount of cytoplasm were noted. These neoplastic cells showed special characteristics, including pleomorphic cells and nuclei, and giant cells. The results indicate cells under mitotic division in some areas. The neoplastic cells in primary tumor appeared in clusters or clumps (Fig. 2A and B). In most areas of samples treated with asafotida, the tumor cells were necrotic and the results showed larger areas of necrosis. Tumor clumps in some areas of tissue were still seen (Fig. 2C).

### 3.3. Microscopic study of the lung

Examination of histological sections of the untreated lungs confirmed the presence of microscopic metastases. Untreated mice exhibited infiltration of neoplastic cells in the lungs and structural destruction of the pulmonary alveoli. The neoplastic cells were characterized by the presence of large hyperchromatic nuclei and small amount of cytoplasm. The metastatic cells in all areas of lungs appeared in clusters or clumps. These cells lining the alveoli and the alveolar septa had several layers (Fig. 3A and B). We investigated the ability of asafotida to modulate breast cancer metastasis to the lung. In microscopic slides of treated lungs, the broad clumps of tumor cells were removed. Tumor cells were seen in some areas of the lung. The alveolar septum had fewer metastatic cells and therefore is less thick (Fig. 3C and D), suggesting that asafotida inhibited the growth of breast cancer metastasis to the lung.

### 3.4. Microscopic study of the liver

Examination of histological sections of the liver confirmed the presence of infiltration of neoplastic cells in the liver. The neoplastic cells were characterized by the presence of large hyperchromatic nuclei and small amount of cytoplasm. The metastatic cells in liver appeared in clusters or clumps. These metastatic cells were wider than the ropes of normal liver hepatocyte plates. The liver lobules were destroyed and central vein and the portal triad were not seen.

A lot of irregular vascular architecture was seen in these samples (Fig. 4A). In treated animals with asafotida, the number of tumor clusters is less than untreated liver. The tumor clusters have lost their integrity and there were necrotic cells, but in some areas, a few neoplastic cells were seen (Fig. 4B).

### 3.5. Microscopic study of breast cancer metastasis to the kidney

Examination of histological sections of the kidney confirmed the presence of infiltration of neoplastic cells in the kidney. The neoplastic cells were characterized by the presence of large hyperchromatic nuclei and small amount of cytoplasm. The metastatic cells in kidney appeared in clusters or small clumps. A large number of tumor cells were similar to renal tubule epithelial cells. They had regular round nuclei and granular eosinophilic cytoplasm. A few number of tumor cells showed clear cytoplasm (clear cells) (Fig. 5A). Most tumor cells in samples treated with asafotida were necrotic and a few number of tumor cells remained in the kidney (Fig. 5B).

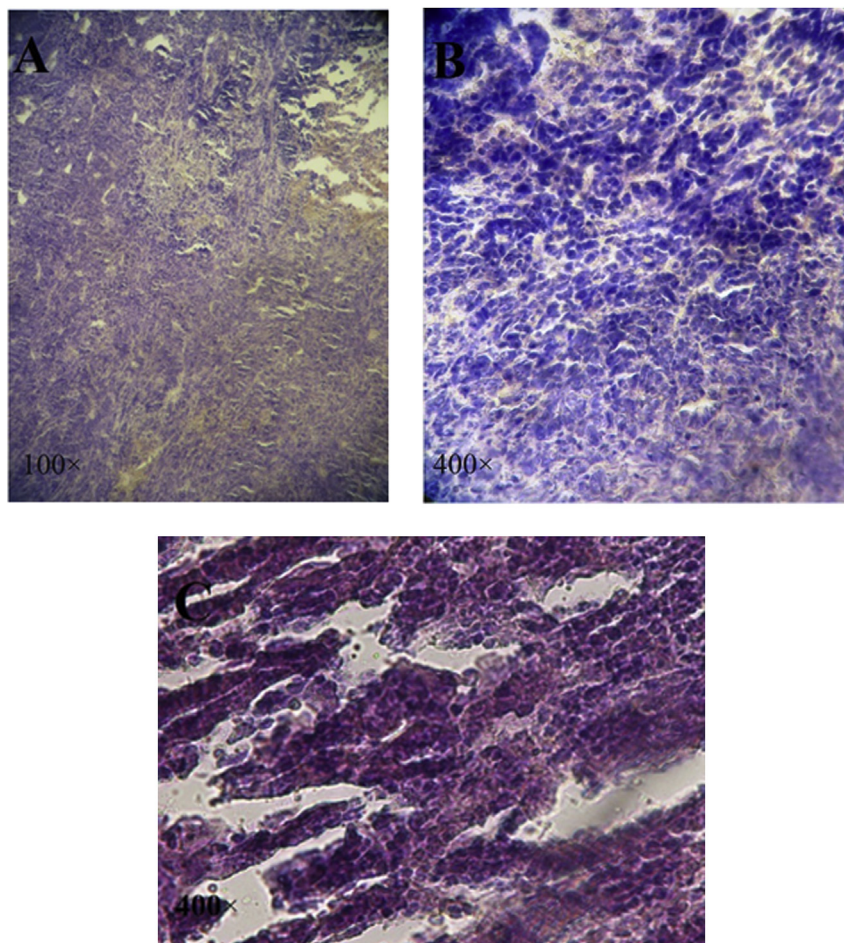
### 3.6. Lipoxigenase inhibitory and radical scavenging activity

The LOX activity was measured as an increase in the absorbance at 234 nm, which reflects the formation of hydroperoxylinoleic acid. The IC<sub>50</sub> asafotida highest inhibitory effect was obtained at 32 µg/ml in this work; our results also showed that the IC<sub>50</sub> of antioxidant activity of asafotida was 109 µg/ml (Table 2).

## 4. Discussion

In the present study, we aimed to evaluate the antitumor and antimetastasis effects of *F. assa foetida* oleo gum resin in a mouse mammary tumor model using the metastasis-specific mouse mammary carcinoma 4T1 cells. The 4T1 breast cancer cells are derived spontaneously from BALB/c mammary carcinoma [23]. These cells closely resemble metastatic breast cancer in human patients and could be used as a suitable model for evaluation of the efficacy of anti-cancer drugs [24]. In this study, our results showed that the weight of the asafotida-treated group decreased for the first 2 weeks after breast cancer induction and started to increase after treatment with asafotida until the end of the experiment. This effect could be due to the ability of asafotida to counteract carcinogenicity/tumor burden, thereby preventing cachexia. Cancer cachexia is defined by loss of muscle mass and fat directly caused by tumor factors or indirectly caused by an aberrant host response to tumor presence [25]. According to the UK Home Office Regulation, weight loss is expected in mice induced with breast cancer and the mice can lose up to 25% of their body weight after 4 weeks of breast cancer induction [26]. In addition, the treatment groups showed a reduction in tumor volume and also the weight of tumors of the asafotida-treated group was lower as compared to the control (Table 1). In a previous study, Bagheri et al. [10] demonstrated the cytotoxic effect of some *Ferula* species on *Artemia salina* as a model for evaluating general cytotoxicity [10]. In different studies, beneficial effects of some *Ferula* species as cancer chemopreventive has been investigated and demonstrated that these species could be as a good source of cancer chemopreventive agents [27]. Asafotida is an oleo-gum-resin obtained from the Iranian endemic medicinal plant, *F. assa foetida* and traditionally it is being used for treatment of various disorders and also have several biological activities such as antioxidant and cancer chemopreventive [28]. In spite of some old evidence about genotoxicity and mutagenicity of asafotida, recent studies have revealed its potential antioxidant, antimutagenicity and cancer chemopreventive activities [11]. In an *in vivo* study, Saleem et al. showed

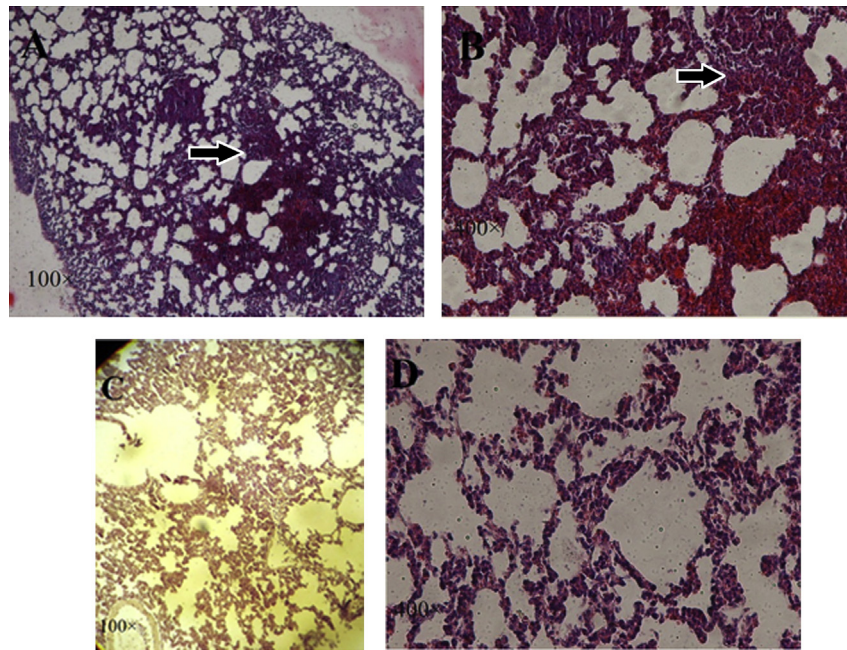




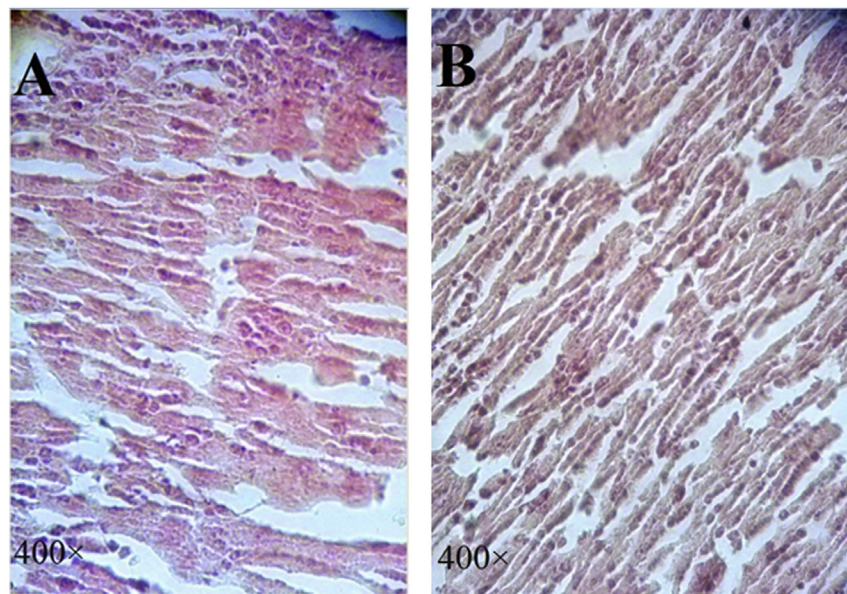
**Fig. 2.** Histology evaluation of primary tumor induced by 4T1, sections from mice untreated and treated with asafetida. H&E staining as observed under a light microscope (100 $\times$ , 400 $\times$ ). In primary tumor, the neoplastic cells were characterized by small amount of cytoplasm. Some cells were under mitotic division (Fig. 2A and B). In sections of treated asafetida breast, numbers of tumor clumps were decreased (Fig. 2C).

that pre-treatment of animals with acetone extract of asafetida could cause the reversal of early events of carcinogenesis [29]. Another study showed that asafetida reduced the multiplicity and size of palpable mammary tumors in Sprague–Dawley rats [11]. Mallikarjuna et al. has reported reduction in the multiplicity and size in mammary tumors in rats following asafetida supplementation [30]. Phytochemical analysis showed that asafetida contains about 40–64% resin, 25% endogenous gum, 10–17% volatile oil and 1.5–10% ash. The resin portion is known to contain farnesiferol, asareninotannols, ferulic acid, umbelliferone, the gum includes glucose, galactose,  $\iota$ -arabinose, rhamnose, glucuronic acid, polysaccharides and glycoproteins, and the volatile fraction contains sulfur-containing compounds, monoterpenes and other volatile terpenoids [11]. Umbelliprenin is one of these components that has been shown to have remarkable cancer chemoprevention *in vitro* and *in vivo* [31]. Moreover, it was reported recently that farnesiferol C from *F. assa-foetida* may be a potential candidate for the treatment of cancer [11]. Phenolic compounds like ferulic acid present in asafetida have been reported to have strong antioxidants and antitumor properties [32]. Ferulic acid has also been reported to inhibit growth of human breast and colon cancer cells [33]. The efficacy of the natural products against cancer has been related with their antioxidant potential to reduce or inhibit free radical mediated damage to cellular macromolecules, such as DNA, lipids

and proteins. Another possible mechanism of action for the active principles could be related to lipoxygenase activity [34]. Interestingly in this study, asafetida showed a remarkable inhibitory activity against lipoxygenase and also antioxidant activity. According to these results, the  $IC_{50}$  of LOX activity of asafetida was calculated as 32  $\mu\text{g/ml}$  (Table 2). In this study we also investigated the treatment effects of asafetida on histopathological and metastasis in lung, liver and kidney in animals treated with asafetida and compared to histological changes in control group. This is the first time that asafetida was demonstrated to be effective not only in decreasing the tumor weight but also in inhibiting lung, liver and kidney metastasis. In most areas of samples treated with asafetida, the tumor cells were necrotic and the results showed larger areas of necrosis. Tumor clumps were still present in some areas of tissue. We also investigated the ability of asafetida to modulate breast cancer metastasis to the lung. In treated lungs, the broad clumps of tumor cells were removed. The alveolar septum had fewer metastatic cells and therefore is less thick. In liver the tumor clusters have lost their integrity and there were necrotic cells, but in some areas, a few neoplastic cells were seen. Histopathological analysis on kidney of treated animals showed that most tumor cells were necrotic. These observations suggest that asafetida under experimental conditions can affect breast carcinogenesis by altering histological changes indicating its anti-carcinogenic potential. The



**Fig. 3.** Histology evaluation of lungs sections from mice untreated and treated with asafoetida. H&E staining as observed under a light microscope (100 $\times$ , 400 $\times$ ). Untreated mice exhibited infiltration of neoplastic cells in the lungs and in the form of clusters or clumps. These cells line the alveoli and the alveolar septa (Fig. 3A and B). In treated lungs, the clumps of tumor cells were removed. The alveolar septum is less thick (Fig. 3C and D). Metastatic cells were marked by arrows.



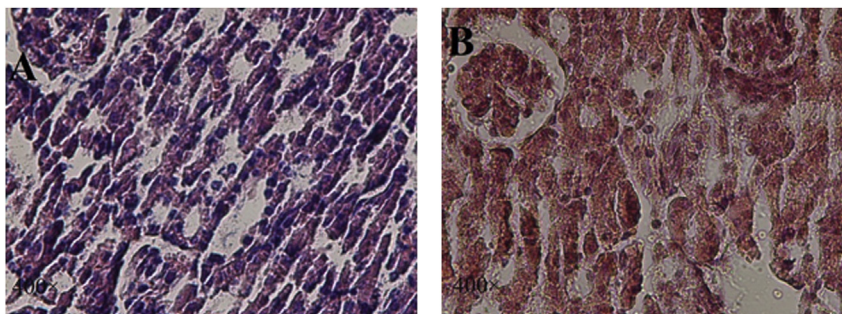
**Fig. 4.** Histology evaluation of liver sections from mice untreated and treated with asafoetida. H&E staining as observed under a light microscope (400 $\times$ ). In untreated liver group, a large number of the tumors cells appeared in clusters or clumps. Central vein and portal triad were not seen and vascular architecture were irregular (Fig. 4A). In sections of the liver treated with asafoetida, there is mild disruption in tumor clusters or clumps (Fig. 4B).

observed modulation in histoarchitecture by asafoetida could be ascribed to its ability to maintain cell structure and integrity either by some direct mechanism or indirectly by scavenging the free radicals. However, studies are required for further exploration with regard to other definitive bioassays including protein expression and documentation of specific molecular markers to establish the exact mechanism for asafoetida mediated cancer chemoprevention.

## 5. Conclusion

The results present the first evidence on the antitumor and antimetastasis effects of asafoetida as demonstrated by *in vivo* tumor weight and metastasis reduction in BALB/c mice bearing breast cancer tumors. More detailed molecular mechanisms, for instance, genomic and proteomic responses underlying asafoetida-induced breast cancer cell apoptotic cell death, and antimetastasis remain to





**Fig. 5.** Histology evaluation of kidney sections from mice untreated and treated with asafoetida. H&E staining as observed under a light microscope (400×). In untreated kidney group, a large number of the tumors cells appeared in clusters or clumps (Fig. 5A). In sections of the kidney treated with asafoetida, a few number of tumor cells remained (Fig. 5B).

**Table 2**

Antioxidant and lipoxygenase inhibitory activities of asafoetida.

DPPH (IC <sub>50</sub> )	Lipoxygenase inhibition (IC <sub>50</sub> )
109 µg/ml	32 µg/ml

be elucidated. Besides, further investigation is needed to determine the clinical efficacy and safety of asafoetida in human subjects with breast cancer metastasis.

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#### Conflict of interest

We certify that no actual or potential conflict of interest related to this article exists.

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